



Efforts towards the synthesis of jatrapholane diterpenes: a biomimetic transannular reaction approach

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Abstract—The construction of the jatrapholane skeleton was accomplished. The key-step of the synthesis is biomimetic transannular C–C bond formation of a lathyrane-type skeleton. © 2001 Elsevier Science Ltd. All rights reserved.

Jatrapholones A and B were isolated from *Euphorbiaceae* (Fig. 1).¹ They possess antitumor activity. From the biogenetic point of view, this 5-6-7-3 fused ring system seems to be derived from a lathyrane skeleton via transannular intramolecular C–C bond formation between the C5 and C12 positions, as shown in Scheme 1. The total synthesis of jatrapholone A was reported only by A. B. Smith III et al. in 1986.²

We are developing a construction method of the skeletons of diterpenes from *Euphorbiaceae* and *Thymeaceae* based on biomimetic skeletal transformation from the lathyrane skeleton as a key intermediate. We previously reported the construction of the lathyrane skeleton. We describe herein the construction of the jatrapholane-type framework by means of transannular radical cyclization.

We first attempted to construct the jatrapholane skeleton via Lewis acid-catalyzed cyclization from the lathyrane-type compound (3) that was reported in our previous paper.³ However, it afforded exclusively the

aldehyde (4),⁴ which was derived from cationic intermediate **5** via C–C bond rearrangement between C7 and C8. (Scheme 2). In the case of the lathyrane intermediate (6) derived from natural euphohelioscopin, the jatrapholane-type compound (7) was produced in moderate yield.⁵ These results suggest that the carbonyl group of the C3 position has a significant role as an intramolecular proton acceptor to abstract a proton from the C6-methyl group to generate the *exo*-methylene group at the C6 position, as shown in Scheme 3.

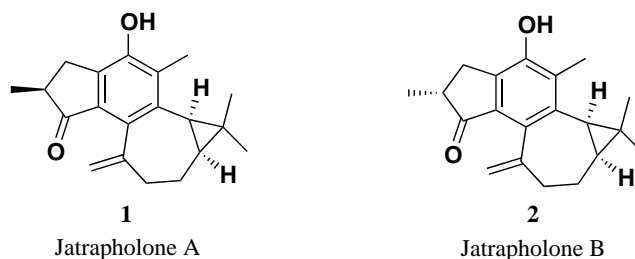
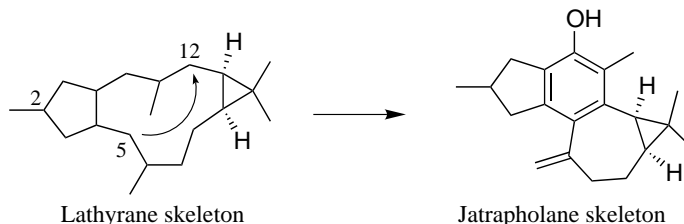


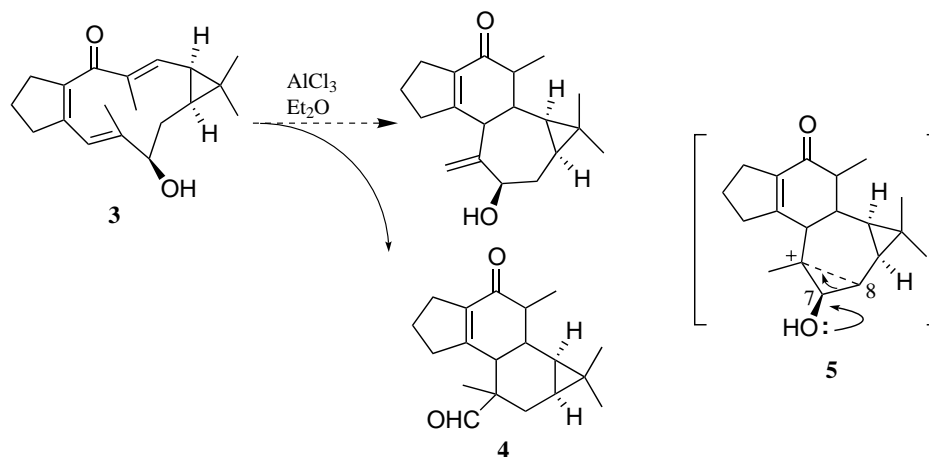
Figure 1.



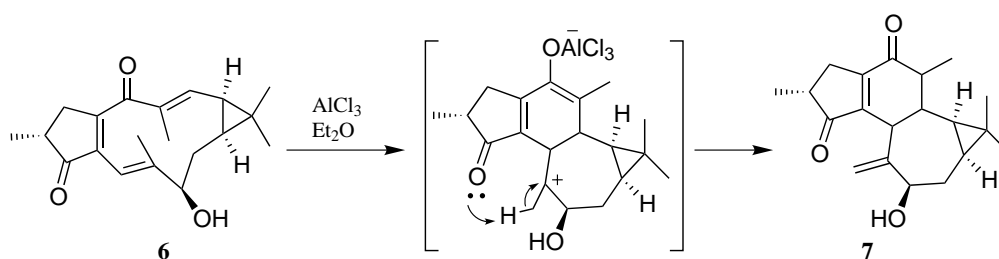
Scheme 1. Biogenetic pathway to the jatrapholane skeleton.

Keywords: cyclization; terpenes and terpenoids; transannular reactions.

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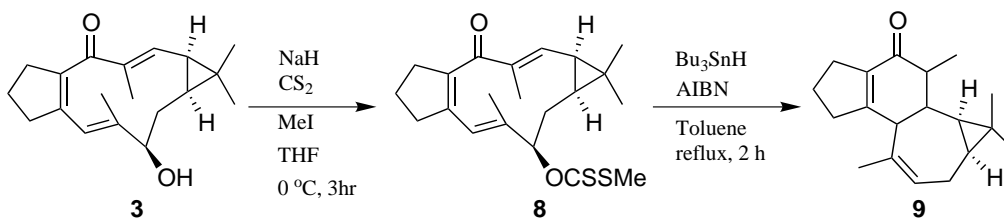
Scheme 2.



Scheme 3.

Therefore, we chose another strategy using a radical cyclization. The C-7 hydroxyl group was converted into xanthate **8**⁴ (71%) by the usual procedure (Scheme 4). In this step the use of thiocarbonyl diimidazol or phenyl chlorothionoformate provided significant amounts of a rearranged dithiocarbonate compound. The xanthate **8** was further treated with tributyltin hydride in the presence of AIBN in refluxed toluene to afford selectively the desired jatrapholane-type compound (**9**)⁶ in 66% yield.

The conformation of **3** was determined to be **3a** by NOE experiments, as shown in Fig. 2. This conformation is compatible with ab initio molecular orbital calculations;⁷ comparing the total energy calculated by the B3LYP method, **3a** ($H_a \cdots H_b = 2.27 \text{ \AA}$) was 11.6852 kJ/mol more stable than **3b**. Herein, a conformational equilibrium of **3** is expected at room temperature, since RHF/6-31G(d) calculations estimated the activation energy of the C5=C6 inversion to be 35.2867 kJ/mol and 22.3824 kJ/mol for **3a** and **3b**, respectively.



Scheme 4.

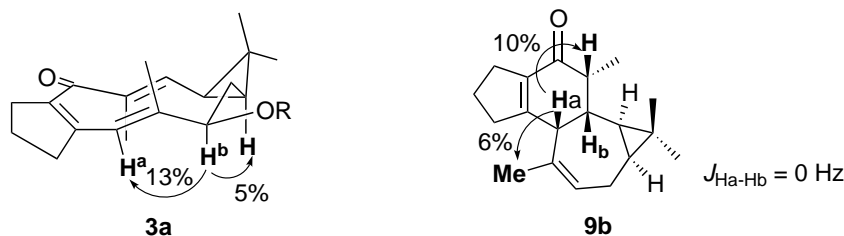
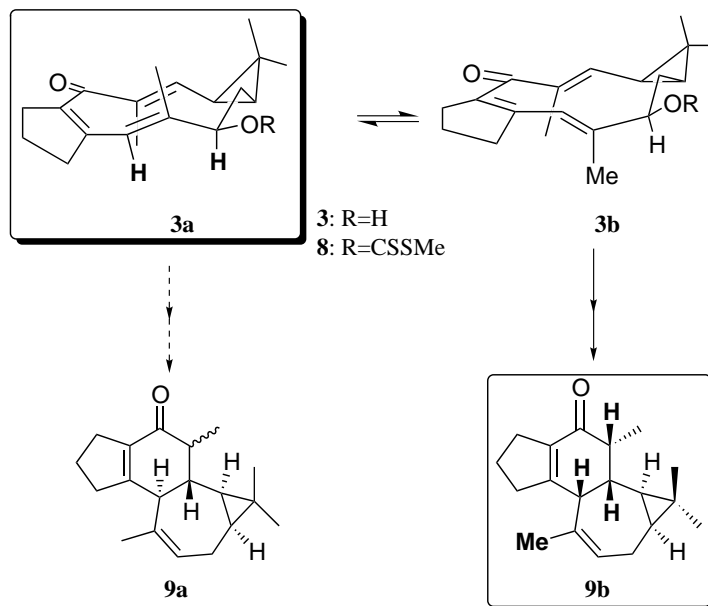


Figure 2.



Scheme 5.

The structure of **9** was expected from the conformation **3a** to be **9a**; however, the NOE experiments of the product indicated that the structure was **9b**, as shown in Fig. 2.

The above reaction consists of two steps: (1) radical formation, and (2) transannular cyclization. Based on the stereostructure of the product (**9b**) and a possible equilibrium between **3a** and **3b**, a plausible mechanism is shown in Scheme 5.

As a result, we accomplished successfully the construction of the jatrapholane-type skeleton from the lathyrane-type skeleton. Work on the total synthesis of jatrapholones A and B is currently under way based on these results.

Acknowledgements

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References

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- The spectral data for the new compounds are in accord with the structures assigned.
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- Spectral data for **9b**: $[\alpha]_D^{19} = -97.9$ (c 0.54, CHCl_3); ^1H NMR (400 MHz, CDCl_3): δ 5.45 (1H, br d, $J=7.5$ Hz), 3.32 (1H, br s), 2.70 (1H, m), 2.6–2.4 (2H, complex), 2.4 (2H, complex), 2.33 (1H, ddd, $J=2.9, 3.5, 10.6$ Hz), 2.20 (1H, ddd, $J=7.5, 8.3, 18.1$ Hz), 1.9 (2H, complex), 1.75 (3H, br), 1.75 (1H, overlapped), 1.07 (3H, d, $J=6.8$ Hz), 0.92 (3H, s), 0.88 (3H, s), 0.67 (1H, ddd, $J=8.3, 8.8, 9.5$ Hz), 0.37 (1H, dd, $J=9.5, 10.6$ Hz); ^{13}C NMR (100 MHz, CDCl_3): δ 199.1, 164.0, 137.0, 133.9, 125.8, 51.9, 49.5, 44.2, 37.1, 29.2, 28.6, 27.7, 24.6, 23.9, 23.8, 22.4, 18.8, 16.7, 12.8; IR (NaCl, film): 2931, 1661, 1448, 1382, 1258 cm^{-1} .
- Ab initio calculations were carried out using the Gaussian 98 series of programs (Gaussian Inc., Pittsburgh, PA, 1998). The starting coordinates of each conformation were generated by inspection of a molecular model, then they were fully optimized using the RHF/6-31G(d) or UHF/6-31G(d) basis set. Successive single-point calculations were performed by B3LYP/6-31G(d) to include electron correlation. The calculations were carried out using a FACOM VPP5000 super computer at Nagoya University Computation Center.